Molybdenum-Catalyzed Olefin Epoxidation: Ligand Effects**

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Abstract: We synthesized substituted pyrazolylpyridine ligands to examine their donor properties by spectroscopic (IR, NMR) and computational (AM 1) methods. The influence of the substitution patterns on spectroscopic and thermodynamic features of molybdenum oxobisperoxo complexes $[(L-L)MoO(O_2)_2]$ (L-L = 2-(1-alkyl-3-pyrazolyl)pyridine/pyrazine) correlates with the activities of the complexes in catalytic olefin epoxidation reactions. This further proof for the relation between the Lewis acidity and the catalytic activity of epoxidation catalysts supports a reaction mechanism in which the peroxo complex activates the oxidizing agent (H_2O_2 , ROOH) instead of directly transferring an oxygen atom from a η^2 -peroxo ligand to the olefin.

Introduction

High-valent molybdenum complexes are known as highly active catalysts for the epoxidation of olefins in the presence of peroxidic reagents like hydrogen peroxide or alkyl hydroperoxides.^[2] During the last two decades, systems with alkoxo, peroxo, acetato, acetylacetonato, halogeno, or oxo ligands have been investigated,^[3] but the chemical constitution of the active species often remained the subject of speculation. In the case of [Mo- $(CO)_6$], for example, which exhibits catalytic activity only after a period of activation, all we currently know about the mechanism is that the low-valent carbonyl complex is oxidized, resulting in the formation of alcoholato complexes, which have not been structurally characterized yet.^[4]

The dearth of structurally well-defined molybdenum(vI) catalysts for olefin epoxidation encouraged us to study Mimountype^[5] molybdenum oxobisperoxo complexes $[(L-L)MoO-(O_2)_2]$, in which L-L is a bidentate 2-(1-alkyl-3-pyrazolyl)pyridine ligand.^[6] From earlier investigations, we knew that these complexes are not subject to ligand exchange reactions under catalytic conditions. Their coordinative stability and their excellent solubility in organic solvents make these peroxo complexes perfect candidates for spectroscopic investigations into the mechanism of catalytic olefin epoxidation. As shown in a previous paper, the oxygen atom transferred to the olefin does not originate from one of the η^2 -peroxo ligands, but from the oxidizing agent.^[7] These results confirm the activation of *t*BuOOH by the Lewis acidic peroxo complexes as a key step in the catalytic cycle.

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above. Pyrazolylpyridines prove to be ideal ligands for that kind of inquiry, since substitution reactions at the pyrazole as well as at the pyridine moiety allow the modification of the donor strength of the chelate system.

Results and Discussion

The chemistry of pyrazole and its derivatives is well established.^[8] A multitude of synthetic routes has been worked out during the last decades, as some members of the pyrazole family play an economically important role in pharmacy and agrochemistry.^[9] In a straightforward synthesis, Claisen condensation of a (mono)alkyl ketone with a carboxylic ester results in the formation of a 1,3-diketone, which can be converted to the corresponding pyrazole by a ring closure reaction with hydrazine. Following this route, Brunner et al. obtained 2-(3(5)pyrazolyl)pyridine (1a) from acetylpyridine, N,N-dimethylformamide dimethylacetal, and hydrazine.^[10] We synthesized the derivatives 1 b-d by variation of either the ketone or the carboxylic ester.^[11] Use of pyrazine carboxylic acid ester instead of the picolinic derivative yielded the corresponding 2-(3(5)-pyrazolyl)pyrazine 1e (Scheme 1).^[12] On the other hand, pyrazoles easily undergo electrophilic substitutions in the 4-position.^[8,13] This feature was applied in the syntheses of the chloro, bromo, and nitro derivatives 1 f-h (Scheme 2).

For a first survey of the influence of the substitution patterns on the electronic situation of the ligands $\mathbf{Ia}-\mathbf{h}$, and thus on their donor properties, we investigated the N-H stretching vibrations of these compounds by IR spectroscopy as well as by computational methods.^[14] Formally, 2-(3-pyrazolyl)pyridines can exist in three tautomeric forms $\mathbf{A}-\mathbf{C}$.^[15] Additionally, a

In this paper we describe the influence of either electron-withdrawing or -donating substituted ligands on the catalytic activities of the molybdenum oxobisperoxo complexes mentioned above. Pyrazolylpyridines prove to be ideal ligands for that kind



Scheme 1. Synthesis of the 2-(3-pyrazolyl)pyridines 1b-e.



1f: Y = Cl; 1g: Y = Br; 1h: Y = NO₂
Scheme 2. Synthesis of the 2-(3-pyrazolyl)pyridines 1f-h.

(small) barrier for the rotation around the pyrazolyl-pyridine bond should be taken into account.^[16] Therefore, six different geometries had to be minimized for each ligand 1a-h in a computational study (Figure 1). Solvent- and temperature-dependent NMR spectra with broad signals, especially for N-H protons, suggest an equilibrium between tautomers/rotamers in solution. For the computational optimization of the geometries and the calculation of thermodynamic data for these species, dihedral angles N2-C-C-N3 of 160° for the tautomers A1, B1, and C1 and of 20° for the corresponding rotamers A2, B2, and C2 were chosen as starting conditions (for atom numbering see Figure 1). As the most important result of our calculations on the systems 1a-h we found that generally B2 is the thermodynamically most stable form (Table 1). It is stabilized by a dipole-dipole interaction between the N2 proton and the lone

Abstract in German: Mehrere substituierte Pyrazolylpyridinliganden wurden synthetisiert und ihre Donoreigenschaften durch spektroskopische Methoden (IR, NMR) und semiempirische Rechnungen (AM1) untersucht. Dabei zeigte sich, daß sich der elektronische Einfluß der Substituenten an den Liganden eindeutig mit spektroskopischen und thermodynamischen Daten der entsprechenden Oxobisperoxomolybdänkomplexe [(L-L)- $MoO(O_2)_2$ (L-L = 2-(1-Alkyl-3-pyrazolyl)pyridin/pyrazin), und deren katalytischer Aktivität bei der Olefinepoxidierung korrelieren läßt. Dies ist ein wichtiger Hinweis auf den Zusammenhang zwischen der Lewis-Acidität von Epoxidierungskatalysatoren und ihrer katalytischen Aktivität. Des weiteren stützen diese Ergebnisse eine über die Aktivierung des Oxidationsmittels durch die Lewis-aciden Peroxo-Komplexe verlaufende Sauerstoffübertragung auf das Olefin, und nicht einen direkten Sauerstofftransfer von einem η^2 -koordinierten Peroxoliganden.



Figure 1. Six isomeric structures of 2-(3-pyrazolyl)pyridine (rotamers/tautomers).

Table 1. Calculated (ΔM 1) relative heats of formation $\Delta\Delta G$ [kJ mol⁻¹] (with respect to **B2**) and torsion angles N2-C-C-N3 [^o] of the six isomeric geometries of 1 a-h.

		1 a	1 b	1 c	1 d	1 e	1 f	1 g	łh
A1	$\Delta\Delta G$	13.49	9.54	8.80	17.27	6.67	15.44	15.34	18.09
	torsion	178	179	179	177	180	149	137	130
B 1	$\Delta\Delta G$	13.49	13.74	13.64	14.39	13.35	12.94	12.61	14.56
	torsion	161	161	164	159	165	148	137	121
C1	$\Delta\Delta G$	75.82	71.34	67.76	62.74	73.84	61.05	62.57	44.22
	torsion	180	180	180	180	180	180	180	163
A 2	$\Delta\Delta G$	17.83	13.85	13.37	22.09	10.88	16.84	15.14	18.57
	torsion	30	30	29	31	26	47	54	50
B 2	$\Delta\Delta G$	0	0	0	0	0	0	0	0
	torsion	0	0	2	1	0	22	30	35
C2	$\Delta\Delta G$	65.37	61.24	57.44	51.48	64.24	60.23	62.63	58.12
	torsion	0	0	0	0	0	0	0	23

pair of N 3. An analogous stabilization could be found for C2 compared with C1 ($1a-e: \Delta G_{(C2, C1)}$ ca. 10 kJmol⁻¹). This particular stabilization can be canceled out if the steric demand of substituents in the 4-position of the pyrazole moiety hampers a coplanar arrangement of the ring systems (1 f,g). In the case of 1h a strong dipole-dipole interaction between the 4-nitro substituent and the N 3 proton is responsible for the stabilization of C1 compared to C2.

In the absence of bulky substituents in the 4-position of the pyrazole ring (ligands 1 a - e), four of the six tautomers/rotamers (A 1, C 1, B 2, C 2) show an almost coplanar arrangement of both rings, while for the two other geometries (B 1, A 2) interplanar angles of $20-30^{\circ}$ are calculated. In the case of these two isomers, a relatively high degree of steric hindrance (H–H interaction) cannot be compensated by π or dipole-dipole interactions. Substitution of the pyrazole proton in the 4-position by more sterically demanding groups (1f-h) leads to nonplanar geometries for A1, B1, A2, and B2, but C1 and C2 remain planar in the case of 1f,g; this can be explained by rather strong dipole-dipole interactions.

However, a coplanar arrangement of the ring systems is necessary for chelating coordination to a metal center. For this reason, beside electronic effects, a substituent at the 4-position of the pyrazole ring should weaken the metal-ligand interaction and therefore increase the Lewis acidity of the metal center.

Calculation of all six N-H stretching frequencies (tautomers/ rotamers A1-C2) for every ligand 1a-h gave a deeper insight into the electronic influence of the substitution patterns on the donor properties of the ligands. Electron-withdrawing substituents, which stabilize the deprotonated (anionic) form and weaken the N-H bond, should shift the N-H absorption to lower wavenumbers. IR experiments in solution resulted for all ligands except 1b in only one N-H absorption. As shown in Table 2 and Figure 2, the experimental data agree quite well

Table 2. Calculated (AM 1, six geometries) and experimental N - H stretching frequencies $[\rm cm^{-1}]$ for the ligands $1\,a{-}h.$

	1 a	1 b	1 e	1 d	1 e	1 f	1 g	1 h
A1	3484.4	3484.2	3475.2	3451.4	3478.7	3476.0	3474.2	3452.3
B 1	3483.1	3483.4	3478.2	3459.0	3479.0	3474.7	3473.4	3451.8
Cl	3440.2	3441.4	3438.6	3424.1	3447.9	3319.6	3254.4	3292.5
A 2	3481.6	3481.8	3473.1	3448.0	3477.7	3473.5	3472.2	3448.1
B 2	3453.9	3451.9	3448.1	3427.0	3452.3	3445.3	3446.9	3425.1
C2	3406.0	3409.4	3404.3	3386.3	3415.5	3392.4	3390.7	3361.5
V. abo	3445	3444	3437	3421	3443	3429	3427	3401
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Figure 2. Correlation of calculated and experimental N–H stretching frequencies of the tautomer B2 (compounds 1a-h) and tautomer A1 of 1b.

with the calculated N-H absorptions of the tautomeric form B2 (R = 0.958). Nevertheless, solvent or concentration effects might influence the experimental values. Together with the thermodynamic data in Table 1, these experiments confirm that B2 is indeed the major tautomer/rotamer in solution. Only in the case of 1b can a second N-H absorption with low intensity be observed. From a thermodynamic point of view, it is obvious that the electron-donating methyl group in the 5-position of the pyrazole ring stabilizes the tautomer A1. The N-H stretching frequency, which was calculated for 1b A1, again matches with the experimental value. The influence of different substituents on v(N-H) could merely be correlated with their Hammett parameters, which are well established only for phenyl derivatives,^[17] but clearly the thermodynamic and spectroscopic properties can be interpreted mainly by inductive effects of the substituents.

Obviously, IR spectroscopy as well as semiempirical calculations are effective tools for the examination of substituent effects in pyrazoles.

We have shown in preceding papers that pyrazolylpyridines with long alkyl side chains are useful ligands for the spectroscopic investigation of seven-coordinate molybdenum peroxo complexes, as they dramatically increase the solubility of these compounds.^[18] Olefin epoxidation reactions can thus be carried out in nonpolar organic solvents like *n*-hexane or toluene, leading to high yields of sensitive epoxides. The ring opening (side-)reaction of the epoxide is suppressed in these systems. Therefore, synthesis of the *N*-octyl derivatives of 1 a-h seemed to be worthwhile. While the *N*-octylated compounds 2a-e could be obtained by reaction of 1a-e with NaH in THF, followed by the addition of octyl iodide, electrophilic substitution of 2a (after *N*-alkylation of 1a) with either chlorine, bromine or nitric acid resulted in higher yields of 2f-h (Scheme 3).



Scheme 3. Synthesis of the N1-octylated 2-(3-pyrazolyl)pyridines 2a-h.

The results of the alkylation reaction do not seem to coincide with the stability of the tautomer **B2**. This contradiction can be elucidated by taking into account the fact that deprotonation of the pyrazolylpyridines with NaH leads to the corresponding anions, which certainly coordinate to the large (with respect to H⁺) sodium cation in a chelating manner. Therefore a nucleophilic attack on the octyl iodide can only proceed by N1 as long as the coordination of the ligand to Na⁺ is strong enough. Only in the case of 1d (CF₃ substituent) could traces of a second compound be observed by GC/MS, showing an almost identical mass spectrum to 2d. It seems that the electron-withdrawing effect of the CF₃ group leads to a weaker metal-ligand interaction and therefore to a (minor) alkylation at N2. As we do not observe any alkylation at N2 in the case of the other 5-substituted derivatives 1 b,c,e, we do not suggest that the regioselectivity of the alkylation is a result of steric hindrance by the ortho substituent pyridine/pyrazine in this position.

The molybdenum peroxo complexes 3a-h can easily be obtained by stirring a methanolic solution of the corresponding

ligand with an excess of hydrated molybdenum oxide ("molybdic acid") in dissolved hydrogen peroxide or by ligand exchange with $[MoO(O_2)_2 \cdot (DMF)_2]$ as source of the molybdenum peroxo unit (Scheme 4).



Scheme 4. Synthesis of the molybdenum peroxo complexes 3a-h.

Owing to the asymmetric nature of the ligand featuring two different coordinating nitrogen donor centers, these peroxo complexes exist in two isomeric forms **A** and **B**, which differ in the orientation of the aromatic rings relative to the axial oxo ligand. In isomer **A** the pyridine ring occupies the second axial position in the same manner as the pyrazole ring in isomer **B**. From spin exchange experiments with **3a** we know that the isomers are in equilibrium in solution (Scheme 5).^[19] Therefore the isomer ratio is not kinetically but thermodynamically deter-



Scheme 5. Equilibrium of the two isomers A and B of 3a-h.

mined. The preferred orientation of the pyridine moiety *trans* to the oxo ligand in **3a** can be explained as a consequence of the better σ donor properties of this fragment compared to pyrazole. Obviously, substitution at both donor systems will have an influence on the equilibrium illustrated in Scheme 5.

Calculations of the donor properties of N2 and N3 were performed for the N1-methylated model ligands 2a'-h'(Scheme 6). We focused on species derived from the type A2



Scheme 6. Molecular structures of the model ligands 2a'-h'. The BH₃ adducts 2a'-h'/N2 and 2a'-h'/N3 were used for calculation of dissociation energies.

(Figure 1), which were calculated in a fixed coplanar orientation of the heteroaromatic rings, since this geometry best matches that of the coordinated ligand in 3a-h. Besides, the results of the calculations depend on the interplanar angle between the pyrazole and the pyridine fragment, because of π interactions between the heteroaromatic rings. In order to determine the donor properties of N2 and N3, dissociation energies of the corresponding BH₃ adducts 2a'-h'/N2 and 2a'-h'/N3(Scheme 6, Table 3) were calculated.

According to the known basicities of 1-methylpyrazole and pyridine^[20] the formation of the BH₃ adduct at the pyridine site is thermodynamically favored; this is confirmed by the differences ($\Delta\Delta G_{N2-N3}$) of the dissociation energies ΔG_{N2} and ΔG_{N3} of the particular adducts. The calculations clearly show the influence of electron-withdrawing and -donating substituents on adduct formation. Additionally, the substitution patterns also determine charge distributions in the molecules, for example, on the BH₃ fragments, and the lengths of the B–N bonds, especially those of the B–N2 bonds (Table 3).

Table 3. Calculated (AM 1) dissociation energies (ΔG_{N2} , ΔG_{N3-N2}) of the BH₃ adducts of $2\mathbf{a}-\mathbf{h}'$ (coplanar geometry), calculated partial charges on the BH₃ fragment, and B-N distances ($d_{\mathbf{a}-\mathbf{N}_2}$) for $2\mathbf{a}-\mathbf{h}'/\mathbf{N}2$ and $2\mathbf{a}-\mathbf{h}'/\mathbf{N}3$.

	Subst.	ΔG _{N2}	ΔG _{№3} Calcd. [kJ mol ⁻¹]	ΔΔG _{N2-N3} Calcd. [kJmol ⁻¹]	2a-h'/N 2 Part. charge on BH ₃ d_{B-N2} [Å]		2a-h'/N 3	
Ligand		Calcd. $[kJ \mod ^{-1}]$					Part. charge on BH ₃	d _{8−№3} [Å]
2a'	CH/4,5-H	58.941	77.516	- 18.576	-0.3167	1.600	- 0.3629	1.590
2 b'	CH/5-CH,	58.666	77.968	- 19.302	-0.3182	1.601	-0.3639	1.590
2c'	CH/5-Ph	56.904	77.595	-20.691	-0.3163	1.601	-0.3632	1.590
2 ď′	CH/5-CF	47.417	75.319	-27.901	-0.2936	1.608	- 0.3596	1.591
2e'	N/5-CH	57.861	74.175	-16.314	-0.3161	1.600	-0.3502	1.584
2 f'	CH/4-CI	51.179	74.111	-22.933	-0.3058	1.610	-0.3622	1.593
2g'	CH/4-Br	50.811	73.848	-23.036	-0.3057	1.610	-0.3617	1.593
2 h'	CH/4-NO ₂	38.952	67.091	-28.140	-0.2851	1.624	-0.3552	1.597

Nevertheless, characterizing the chelate ligands 2a-h as a geometrically fixed arrangement of two independent donor centers would not be consistent with the results of our calculations. Electron-withdrawing groups, such as on the pyrazole moiety, also destabilize the BH₃-N 3 adduct. This can be explained in terms of the π interactions between both ring systems being coplanar (as in chelate complexes). These facts have to be kept in mind for the following discussion of the particular Lewis acidities of the peroxo complexes 3a-h.

However, the equilibrium constants for the reactions $\mathbf{A} \rightleftharpoons \mathbf{B}$ (of $3\mathbf{a}-\mathbf{h}$) should only depend on the relative donor strengths of N3 and N2. An almost linear relation (R = 0.955) can be obtained between the calculated $\Delta\Delta G_{N2-N3}$ and ΔG_{Mo} ($\Delta G_{Mo} =$ $- R T \ln([A]/[B])$, determined by ¹H NMR spectroscopic measurements (Table 4, Figure 3). Consequently, the combination of spectroscopic data with theoretical calculations allows a qualitative description of some characteristics of these catalytically relevant transition metal complexes.

Table 4. Calculated dissociation energies $\Delta\Delta G_{N3-N2}$ and equilibrium constants $K_{N3/N2}$ of the BH₃ adducts of $2\mathbf{a'}-\mathbf{h'}$, and experimental (NMR) isomer ratios (A:B) and energy differences ΔG_{M0} of the peroxo complexes $3\mathbf{a}-\mathbf{h}$.

Ligand	Subst.	$K_{N3/N2}$ calcd.	$\Delta\Delta G_{N2-N3}$ caled. [kJ mol ⁻¹]	Isomer ratio A:B	ΔG_{Mo} exp. [kJ mol ⁻¹]
2 a'	CH/4,5-H	1803	-18.576	1.71	1.329
2 b′	CH/5-CH ₃	2417	-19.302	0.98	0.050
2c′	CH/5-Ph	4236	- 20.691	1.42	0.869
2 d′	CH/5-CF ₃	77764	-27.901	28.9	8.334
2 e'	N/5-CH	724	-16.314	0.084	6.137
2 f'	CH/4-Cl	10470	-22.933	7.05	4.839
2 g'	CH/4-Br	10914	-23.036	7.86	5.108
2h′	CH/4-NO ₂	85639	-28.140	> 50	< 9.7



Figure 3. Correlation of calculated (2a'-h') and experimental $(3a-h) \Delta G$ values.

In the case of the most electron-deficient pyrazole moiety (3h, nitro substituted), isomer B could not be observed at all. In contrast, complex 3e shows an "inverse" isomer ratio, where the methyl substituent increases the electron density of the five-ring system, while the exchange of pyridine for pyrazine leads to the opposite effect for the six-ring heteroaromatic fragment.

Not only are the spectroscopic properties of the peroxo complexes 3a-h influenced by their specific ligand substitution pattern, but also their epoxidation activities should depend on the electronic structure of the chelating ligand. As a result of mechanistic investigations we know that in the case of these sevencoordinate complexes, the oxygen atom transferred to the olefin originates from the oxidizing agent *t*BuOOH and not from a η^2 -O₂ ligand of the peroxo complexes.^[7, 21] The activation of the hydroperoxide by a high-valent metal center is achieved through an interaction between the Lewis acidic metal complex and the Lewis basic hydroperoxide,^[22] promising highly active complexes bearing (chelate) ligands with electron-withdrawing substituents.

In the first step of the proposed reaction mechanism, the hydroperoxide coordinates to the peroxo complex, followed by a proton transfer reaction (Scheme 7). η^2 -coordination of the hydroperoxide to the Lewis acidic metal center activates the oxidizing agent towards electrophilic attack at the olefinic double bond. Reprotonation of the resulting alcoholato ligand and dissociation of the alcohol regenerates the ligand sphere of the oxobisperoxo complexes.



Scheme 7. Proposed mechanism for the catalytic olefin epoxidation by seven-coordinate molybdenum oxobisperoxo complexes of type 3a-h.

In the case of our molybdenum oxobisperoxo complexes $3\mathbf{a}-\mathbf{h}$, the situation is even more complicated, because they exist in two isomeric forms in equilibrium at room temperature. It must be considered that all the other species involved in the mechanism can also show this structural feature. At present, we do not know anything about the equilibria between these isomers and their particular contribution to the overall activity of the catalytic system. We have just started a detailed NMR study on the coordination chemistry and ligand fluxionality of the complexes $3\mathbf{a}-\mathbf{h}$.

From catalytic epoxidations carried out at various temperatures we found a temperature-dependent activation period wherein the actual catalytically active species is formed. This catalyst species is therefore not identical with the oxobisperoxo molybdenum complex. We assume it to be a hydroperoxo alkylperoxo molybdenum complex as suggested in Scheme 7, but spectroscopic or structural evidence for this species is still lacking. At reaction temperatures above 50 °C the activation periods of the complexes 3a-h are too short to be observed by GC analysis, indicating that the reaction rates determining the first equilibrium of the mechanism (formation of ROOMoOOH from $Mo(\eta^2-O_2)$ and ROOH) are high. Under these conditions the oxygen transfer from the active species to the olefin is the rate-determining step at the beginning of the reaction. For a qualitative description of the actual epoxidation activities of complexes 3a-h, the initial turnover frequencies (TOF, Table 5) were calculated. Cyclooctene was used as the standard system.

Table 5. Experimental initial TOFs for the catalytic epoxidation of cyclooctene with the oxobisperoxo complexes 3a-h (for conditions see Experimental Part), and their relative reactivities with respect to 3a.

Complex	TOF $[h^{-1}]$	Rel. Activity
	3420	1.00
3 b	3160	0.92
3c	4040	1.18
3 d	5710	1.67
3e	4940	1.45
3f	4930	1.44
3g	5200	1.52
3h	6470	1.89

In addition to the spectroscopic and thermodynamic features, the substitution pattern of the chelating ligand also controls the catalytic activity of the peroxo complexes 3a-h. Electron-withdrawing groups, which elevate the Lewis acidity of the catalyst, are responsible for an increase in activity. In comparison to **3a**, the nitro substituent of 3h, for example, increases the reactivity of this compound by a factor of almost two, while the methyl substituent in 3b decreases its reactivity to 90%. Interestingly enough, substitution of pyrazine for the pyridine system overrules the influence of the methyl group at the pyrazole ring and leads to a higher reactivity of **3e**. Both the linear plot of the ΔG values and the logarithmic plot of the isomer ratios of 3a-h (derived from NMR spectroscopy) against the reactivity data (see Figure 4) result in a linear correlation (R = 0.979) for the pyrazolylpyridine complexes 3a-d and 3f-g. Obviously, the data for the pyrazine derivative 3e cannot be correlated this way.



Figure 4. Logarithmic correlation of the initial TOFs and the isomer ratios of the peroxo complexes **3a-h**.

Thus the isomer ratios and the corresponding ΔG values for the complexes correlate with the donor properties of the particular ligands. Consequently, the electronic properties of the ligands determine the activities of the complexes $3\mathbf{a}-\mathbf{h}$. Steric effects should only play a minor role.

Conclusion

With a combination of spectroscopic, computational, and kinetic methods, we are now able to quantify the electronic influence of ligands on the activity of epoxidation catalysts. On the basis of the results, rules for the ligand design of epoxidation catalysts can be established. As mentioned in previous papers, coordinative stability of the catalytically active complexes and stability against oxidative degradation of the ligand system could be achieved by application of neutral chelate ligands bearing two nitrogen donor centers as parts of aromatic ring systems. The most simple and familiar ligand fulfilling these demands is 2,2'bipyridine. Owing to the low solubility of bipyridine complexes of high-valent metal centers in organic solvents, a further structural feature for an optimal ligand (for catalysis in homogeneous systems) is required: alkyl side chains lead to increased solubility of the catalysts. Synthetic considerations directed us towards the pyrazolylpyridine system, the donor properties of which can easily be optimized. As shown above, exchange of the pyridine for a less electron-rich pyrazine moiety results in increased activity in olefin epoxidation. Analogously, the introduction of a second pyrazolyl fragment instead of the pyridyl ring will improve the catalytic activity and allow further introduction of electron-withdrawing substituents. Our future research will concentrate on the synthesis of electron-deficient bispyrazolyl ligands in which the nitrogen bears -CH₂COOR moieties instead of electron-rich alkyl groups. These substituents have already proved to be ideal side chains with which to increase the catalytic activity of our peroxo complexes.^[6]

Experimental Section

General information: Ligands were prepared under nitrogen atmosphere; solvents were dried and distilled before use. 2-(3(5)-pyrazolyl)pyridine (**1**a),^[10] 2-(1-octyl-3-pyrazolyl)pyridine (**2**a),^{(6]} (1-octyl-3-pyrazolyl)pyridineoxodiperoxomolybdenum(vi) (**3**a),^[6] and [MoO(O₂)₂ · (DMF)₂]^[5] were synthesized as described in the literature. The NMR (Bruker DPX 400), mass (gas chromatograph Hewlett – Packard HP 5890 coupled with a mass-selective detector HP 5970, Finnigan MAT 90), and infrared spectra (Perkin – Elmer 1600 series FTIR), and all elemental analyses were carried out at the Anorganisch-chemisches Institut der Technischen Universität München. The

samples from the catalytic epoxidation of cyclooctene were analyzed by GC (Hewlett– Packard GC HP5890 Series II, FI detector, internal standard: dibutyl ether). The numbering scheme for NMR spectra assignments is given in Scheme 8.

General procedure for the syntheses of ligands 1b-1e by Claisen condensation and ring-closing reaction of the resulting 1,3-dione with hydrazine: Sodium (2.30 g, 100 mmol) was dissolved in ethanol (100 mL), the solvent



was removed in vacuo, and the resulting KOEt was dissolved in THF (100 mL). To this solution, the ketone (50.0 mmol) was added, followed by the ester (50.0 mmol), both dissolved in THF (50 mL). After heating under reflux for 6 h, the solvent was removed again and the yellow-to-brownish solid was dissolved in water (50 mL). The aqueous solution was neutralized with acetic acid and extracted with diethyl ether (2×50 mL). The organic phase was dried over MgSO₄ and the solvent was removed in vacuo to yield the 1,3-dione. A solution of the crude 1,3-dione in ethanol (100 mL) was treated with an excess of hydrazine (80% in water) and refluxed for 5 h. The solution was evaporated and the resulting solid was purified by sublimation in vacuo.

2-(5-Methylpyrazol-3-yl)pyridine (1 b):^[11a] From acetone and *o*-picolinic acid methyl ester. Yield: 47 %, pale yellow microcrystals. M.p.: 115 °C; IR (KBr): $\tilde{v} = 3202 \text{ s m}^{-1}$, 3127 s, 3094 m, 3041 m, 2981 m, 2940 m, 2870 s, 1600 vs, 1564 s, 1505 s, 1470 s, 1439 s, 1408 s, 1372 m, 1275 m, 1250 m, 1166 m, 1144 m, 1085 s, 1050 m, 1023 m, 1000 s, 966 m, 958 m, 884 m, 854 s, 778 vs, 737 s, 690 m, 624 s, 513 m; IR (CH₂Cl₂): $\tilde{v} = 3511 \text{ w cm}^{-1}$, 3444 s, $(2 \times \text{NH})$; ¹H NMR (400.13 MHz, 25 C, CDCl₃): $\delta = 8.60$ (d, ³J_{10.11} = 4.8 Hz, 11-H), 7.66 (m, 8-H, 9-H), 7.16 (ddd, ³J_{9.10} = 6.5 Hz, ⁴J_{8.10} = 1.3 Hz, 10-H), 6.54 (br, 4-H), 2.31 (s, 3H, CH₃), NH not observed; ¹³C{¹H} NMR (100.63 MHz, 25 °C, CDCl₃): $\delta = 149.3$ (C-7), 148.8 (C-11), 145.9 (C-3), 145.5 (C-5), 136.2 (C-9), 121.9 (C-10), 119.4 (C-8), 102.4 (C-4), 12.0 (CH₃); MS (EI, 70 eV): *m/z* (%) = 159 (100) [*M*⁺], 130 (68) [*M*⁺ - HN₂], 104 (6) [*M*⁺ - C₂N₂H₄], 103 (8) [*M*⁺ - C₂N₂H₅], 78 (7) [C₅H₄N⁺]; C₉H₉N₃ (159.2): caled C 67.90, H 5.70, N 26.40; found C 68.01, H 5.90, N 26.48.

2-(5-Phenylpyrazol-3-yl)pyridine (1 c):^[11b] From acetophenone and *o*-picolinic acid methyl ester. Yield: 50%; m.p.: 162 °C; IR (KBr): $\tilde{v} = 3244 \text{ s cm}^{-1}$, 3061 m, 2862 w, 2923 m, 2853 w, 1597 m, 1567 m, 1470 s, 1450 s, 1385 w, 1314 w, 1297 w, 1175 m, 1076 m, 1047 m, 995 m, 972 m, 957 m, 912 w, 802 w, 780 m, 759 vs, 734 w, 720 w, 691 w, 658 w, 620 w, 517 w, 508 w, 486; IR (CH₂Cl₂): $\tilde{v} = 3437 \text{ s cm}^{-1}$ (NH); ¹H NMR (400.13 MHz, 25 °C, CDCl₃): $\delta = 8.69$ (d, ³ $J_{10,11} = 4.5 \text{ Hz}$, 11-H), 7.87 (m, 8-H, 9-H), 7.74 (d, ³ $J_{0,m} = 3.5 \text{ Hz}$, o-H), 7.42 (t, ³ $J_{m,p} = 7.5 \text{ Hz}$, m-H), 7.32 (t, p-H), 7.32 (dd, ³ $J_{1,10} = 7.5 \text{ Hz}$, 10-H), 7.05 (s, 4-H), NH not obs.; ¹³C{¹H} NMR (100.63 MHz, 25 °C, CDCl₃): $\delta = 151.6$ (C-7), 149.5 (C-11), 148.6 (C-3), 144.6 (C-5), 137.0 (C-9), 132.6 (C-*i*), 128.7 (C-*o*), 128.0 (C-*p*), 125.7 (C-*m*), 122.9 (C-10), 120.1 (C-8). 100.4 (C-4); MS (EI, 70 eV): m/z (%) = 221 (100) $\{M^+]$, 192 (43) $[M^+ - N_2H]$, 191 (20) $[M^+ - N_2H_2]$, 78 (6) $[C_5H_4N^+]$; $C_{14H_1N_3}$ (231.3): calcd C 76.00, H 5.01, N 18.99; found C 75.40, H 5.04, N 18.83.

2-(5-Trifluoromethylpyrazol-3-yl)pyridine (1d):^[11c] From 2-acetylpyridine and trifluoroacetic acid ethyl ester. Yield: 52% of the monohydrate, pale yellow microcrystals. M.p.: 130 °C; IR (KBr): $\tilde{v} = 3297 \,\mathrm{s} \,\mathrm{cm}^{-1}$, 3092 m, 2854 m, 2707 m, 1598 m, 1585 s, 1567 m, 1481 m, 1440 m, 1414 m, 1361 s, 1342 m, 1288 m, 1257 s, 1178 vs, 1137 s (2 × CF), 1099 w, 1078 w, 1029 s, 1013m, 996m, 967w, 899m, 852m, 782s, 748m; IR (CH₂Cl₂): $\tilde{v} = 3421 \text{ s cm}^{-1}$ (NH); ¹H NMR (400.13 MHz, 25 °C, [D₆]acetone): $\delta = 8.55$ $(ddd, {}^{3}J_{10,11} = 4.8 \text{ Hz}, {}^{4}J_{9,11} = 1.7 \text{ Hz}, {}^{5}J_{8,11} = 0.9 \text{ Hz}, 11\text{-H}), 7.94 (dt, 3.11)$ ${}^{3}J_{8,9} = 7.1$ Hz, ${}^{4}J_{8,10} = 1.1$ Hz, 8-H), 7.77 (ddd, ${}^{3}J_{9,10} = 7.6$ Hz, 9-H), 7.68 (br, 4-H), 7.30 (ddd, 10-H), 6.51 (br, N–H), 3.50 (s, 2H, H_2O); ¹³C{¹H} NMR (100.63 MHz, 25 °C, [D₆]acetone): $\delta = 152.3$ (C-7), 150.6 (C-3), 149.9 (C-11), 137.1 (C-9, C-4), 125.0 (q, ${}^{1}J_{C,F} = 271.5 \text{ Hz}, \text{ CF}_{3}$), 124.1 (C-10), 120.7 (C-8), 92.6 (q, ${}^{2}J_{C,F} = 31.1$ Hz, C-5); MS (EI, 70 eV): m/z (%) = 213 (100) $[M^+]$, 194 (10) $[M^+ - F]$, 165 (52) $[M^+ - HF - N_2]$, 145 (10) $[M^+ - 2 \text{HF} - N_2], 138 (15) [M^+ - \text{HFCN}_3], 136 (6) [C_4 \text{HF}_3 N_2^+], 78 (21)$ [C₅H₄N⁺]; C₉H₆F₃N₃·H₂O (231.2): calcd C 46.76, H 3.49, N 18.18: found C 46.57, H 3.46, N 17.97.

2-(5-Methylpyrazol-3-yl)pyrazine (1e): From acetone and pyrazinecarboxylic acid ethyl ester. Yield: 44.1%, pale yellow microcrystals. M.p.: 118 °C; IR (KBr): $\hat{v} = 31898 \text{ cm}^{-1}$, 3139 s, 3113 m, 3043 w, 2955 m, 2924 s, 2876 m, 2853 m, 1587 m, 1524 s, 1506 m, 1467 m, 1455 m, 1419 m, 1376 m, 1275 w, 1261 w, 1160 m, 1147 m, 1089 m, 1028 s, 1018 s, 969 m, 847 m, 802 m, 669 m, 410 m; IR (CH₂Cl₂): $\hat{v} = 3443 \text{ s cm}^{-1}$ (NH); ¹H NMR (400.13 MHz. 25 °C, [D₆lacetone): $\delta = 12.05$ (br, N–H), 9.04 (s, 8-H), 8.42 (d, ³J_{10,11} = 2.5 Hz, 10-H), 8.34 (d, 11-H), 6.57 (s, 4-H), 2.23 (s, CH₃); ¹³C[¹H} NMR (100.63 MHz, 25 °C, [D₆lacetone): $\delta = 145.8$ (C-7, C-3), 144.8 (C-11, C-10), 143.4 (C-8, C-5), 105.0 (C-4), 12.16 (s, CH₃); MS (EI, 70 eV): *m/z* (%): 160 (100) [*M*⁺], 131 (64) [*M*⁺ – HN₂], 106 (30) [*M*⁺ – C₃H₄N], 77 (21) [C₅H₃N⁺], 66 (7) [C₄H₄N⁺], 51 (46) [C₃HN⁺]; C₈H₈N₄ (160.2): caled C 59.99, H 5.03, N 34.98; found C 59.57, H 5.24, N 35.16.

2-(4-Chloropyrazol-3-yl)pyridine (1f): Compound **1a** (1.45 g, 10.0 mmol) was dissolved in 6% HCl (100 mL) and stirred at room temperature while a saturated solution of chlorine in water (400 mL) was added in small portions over a period of 3 h. After another 2 h, ammonium acetate (15 g) was added and the pale yellow solution was neutralized with KOH. The resulting colorless precipitate was filtered off, washed with water, dried in vacuo, and purified by sublimation. Yield: 1.61 g (91%), white crystals. M.p.: 152 °C; 1R (KBr): $\tilde{\nu} = 3414$ m cm⁻¹, 3168 vs, 3115 s, 3045 m, 2984 m, 2950 m, 1598 s, 1572 m, 1560 m, 1482 s, 1469 m, 1411 m, 1327 m, 1319 m, 1215 w, 1152 m,

1103 m, 1092 m (C–Cl), 1059 m, 1015 w, 989 m, 928 s, 841 m, 786 s, 742 m, 672 s, 626 m, 555 w; IR (CH₂Cl₂): $\tilde{\nu} = 3429 \text{ s cm}^{-1}$ (N-H); ⁻¹H NMR (400.13 MHz, 25 °C, CDCl₃ + [D₆]DMSO): $\delta = 8.56$ (d, ³ $J_{10,11} = 4.5$ Hz, 11-H), 7.94 (d, ³ $J_{8,9} = 8.0$ Hz, 8-H), 7.72 (dt, ³ $J_{9,10} = 8.0$ Hz, ⁴ $J_{9,11} = 1.5$ Hz, 9-H), 7.49 (s, 1 H, 5-H), 7.20 (dd, 10-H), NH not det.; ¹³C{¹H} NMR (100.63 MHz, 25 °C, CDCl₃ + [D₆]DMSO): $\delta = 148.3$ (C-11), 135.8 (C-9), 121.9 (C-10), 120.0 (C-8), 106.7 (C-4), broad (C-3), (C-5), (C-7); MS (EI, 70 eV, ³⁵CI): m/z (%): 179 (100) [M^+], 151 (15) [$M^+ - N_2$], 144 (3) [$M^+ -$ Cl], 116 (40) [$M^+ - N_2 -$ Cl], 89 (33) [C₂H₂N₂Cl⁺], 78 (14) [C₃H₄N⁺]; C₈H₆ClN₃ (179.6): calcd C 53.50, H 3.37, N 23.40; found C 53.49, H 3.32, N 23.78.

2-(4-Bromopyrazol-3-yl)pyridine (1g): Compound 1a (1.45 g, 10.0 mmol) was dissolved in 20% acetic acid (100 mL) and stirred at room temperature while bromine (1.59 g, 10.0 mmol), dissolved in glacial acetic acid (10 mL), was added over 15 min. The resulting pale yellow solution was neutralized with KOH. The colorless precipitate was filtered off, washed with water, dried in vacuo, and purified by sublimation. Yield: 1.99 g (89%), pale yellow needles. M.p.: 148 °C; IR (KBr): $\tilde{v} = 3417 \,\mathrm{m} \,\mathrm{cm}^{-1}$, 3172 vs, 3110 s, 3038 m, 2978 w, 2945 w, 2850 w, 1595 s, 1572 m, 1555 m, 1482 m, 1459 m, 1417 w, 1407 m, 1316 m, 1214 w, 1151 w, 1105 m, 1058 m, 1006 m (C-Br), 984 m, 928 s, 844 m, 786 s, 742 m, 670 s, 626 w, 516 w; IR (CH₂Cl₂): $\tilde{v} = 3427$ s cm⁻¹ (N-H); ¹H NMR (400.13 MHz, 25 °C, CDCl₃): δ = 12.33 (br, N-H), 8.66 (d, ${}^{3}J_{10,11} = 4.5$ Hz, 11-H), 8.28 (d, ${}^{3}J_{8,9} = 8.0$ Hz, 8-H), 7.80 (dt, ${}^{3}J_{9,10} = 8.0$ Hz, ${}^{4}J_{9,11} = 1.5$ Hz, 9-H), 7.63 (s, 1 H, 5-H), 7.29 (dd, 10-H); ¹³C{¹H} NMR (100.63 MHz, 25 °C, CDCl₂): $\delta = 149.4$ (C-11), 146.9 (C-7), 142.1 (C-5), 138.0 (C-3), 137.1 (C-9), 123.4 (C-10), 120.6 (C-8), 92.3 (C-4); MS (EI, 70 eV, ⁷⁹Br): m/z (%) = 223 (75) [M^+], 197 (5) [$M^+ - N_2$], 144 (10) $[M^+ - Br]$, 116 (100) $[M^+ - Br - N_2]$, 89 (76) $[C_6H_3N^+]$, 78 (56) [C₅H₄N⁺]; C₈H₆BrN₃ (224.1): calcd C 42.88, H 2.70, N 18.76; found C 42.63, H 2.70, N 18.73.

2-(4-Nitropyrazol-3-yl)pyridine (1h): Compound 1a (1.45 g, 10.0 mmol) was dissolved in 80% H₂SO₄ (10 mL) at 0 °C. To this solution, a mixture of 65% HNO3 (5 mL) and 80% H2SO4 (5 mL) was added dropwise over a period of 30 min. After the reaction mixture had been heated to 90 °C for 4 h, it was poured over ice (300 g) and carefully neutralized with conc. K₂CO₃. The resulting colorless precipitate was filtered off, washed with water, dried in vacuo, and purified by sublimation. Yield: 1.18 g (62%), white microcrystals. M.p.: 167 °C; IR (KBr): $\tilde{v} = 3169 \,\mathrm{m} \,\mathrm{cm}^{-1}$, 3139 s, 3128 s, 3103 m, 3038 m, 2968 m, 2921 m, 2887 w, 2799 w, 1594 m, 1564 m, 1543 m, 1500 s $(N=O)_{as}$, 1488s, 1464s, 1422s, 1386s, 1354m, 1330vs $(N=O)_{sym}$, 1290w, 1252 w, 1216 m, 1178 w, 1161 m, 1095 m, 1065 m, 1030 w, 994 m, 923 m, 881 m. 849 w, 832 m, 791 s, 757 s, 679 w, 628 w; 1R (CH $_2$ Cl $_2$): $\tilde{v} = 3401$ s cm $^{-1}$ (N – H); ¹H NMR (400.13 MHz, 25 °C, $[D_6]$ acetone): $\delta = 13.05$ (br, N–H), 8.76 (d, ${}^{3}J_{10,11} = 4.2$ Hz, 11-H), 8.45 (br, 5-H), 8.08 (br, 8-H), 7.99 (t, ${}^{3}J_{8,9} = {}^{3}J_{9,10} = 7.3$ Hz, 9-H), 7.55 (t, 10-H); ${}^{13}C{}^{1}H{}$ NMR (100.63 MHz, 25° C, [D₆]acetone): $\delta = 149.4$ (C-7), 149.3 (C-11), 149.1 (C-3), 136.8 (C-9), 132.5 (C-5), 124.4 (C-10), 124.3 (C-8), 123.6 (C-4); MS (EI, 70 eV): m/z $(\%) = 190(55)[M^+], 160(2)[M^+ - NO], 116(37)[M^+ - C_5H_4N], 105(54)$ $[C_6H_5N_2^+]$, 89 (100) $[C_6H_3N^+]$, 78 (57) $[C_5H_4N^+]$; $C_8H_6N_4O_2$ (190.2): calcd C 50.53, H 3.18, N 29.46; found C 50.37, H 3.25, N 29.66.

General procedure for the syntheses of the octyl derivatives 2b-e by alkylation of 1b-e with octyl iodide: NaH (0.24 g, 10 mmol) was suspended in THF (50 mL). To this suspension, the ligands 1b-e (10 mmol) were added and the mixture was stirred until the evolution of hydrogen stopped. Then $C_8H_{17}I$ (2.40 g, 10 mmol) was added and the resulting solution was refluxed for 24 h. After removing the solvent in vacuo, the product was extracted from the oily residue with boiling hexane. The ligands 2b-e were obtained in 60-80%yield as oils with sufficient purity (GC, NMR) for the syntheses of the peroxo complexes.

2-(5-Methyl-1-octylpyrazol-3-yl)pyridine (2b): ¹H NMR (400.13 MHz, 25 °C. CDCl₃): $\delta = 8.50$ (dd, ³ $J_{10,11} = 4.5$ Hz, ⁴ $J_{9,11} = 2.0$ Hz, 11-H), 7.79 (dd, ³ $J_{8,9} = 8.0$ Hz, ⁴ $J_{8,10} = 1.0$ Hz, 8-H), 7.60 (dt, ³ $J_{9,10} = 8.0$ Hz, 9-H), 7.06 (ddd, 10-H). 6.53 (s, 4-H), 3.98 (t, ³ $J_{\text{H,H}} = 8.0$ Hz, NCH₂), 2.24 (s, C5-CH₃), 1.77 (br, NCH₂CH₂), 1.22–1.18 (br, 10H, CH₂), 0.80 (t, ³ $J_{\text{H,H}} = 6.5$ Hz, CH₂CH₃); ¹³C{¹H} NMR (100.63 MHz, 25 °C, CDCl₃): $\delta = 152.4$ (C-7), 149.8 (C-3), 149.2 (C-11), 139.2 (C-5), 136.3 (C-9), 121.9 (C-10), 119.7 (C-8), 103.7 (C-4), 49.3 (NCH₂), 31.5–22.4 (CH₂), 13.9 (CH₂CH₃), 11.1 (C5-CH₃); MS (EI, 70 eV): m/z (%) = 271 (20) [M^+], 256

 $\begin{array}{l} (5) \ [M^+ - \mathrm{CH_3}], \ 242 \ (9) \ [M^+ - \mathrm{C_2H_5}], \ 228 \ (26) \ [M^+ - \mathrm{C_3H_7}], \ 214 \ (21) \\ [M^+ - \mathrm{C_4H_9}], \ 200 \ (6) \ [M^+ - \mathrm{C_5H_{11}}], \ 186 \ (16) \ [M^+ - \mathrm{C_6H_{13}}], \ 172 \ (100) \\ [M^+ - \mathrm{C_7H_{15}}], \ 159 \ (35) \ [M^+ - \mathrm{C_8H_{16}}], \ 145 \ (6) \ [M^+ - \mathrm{C_8H_{16}} - \mathrm{CH_3}], \ 130 \\ (19) \ [M^+ - \mathrm{C_8H_{17}N_2}], \ \ 117 \ \ (10) \ \ [M^+ - \mathrm{C_8H_{16}} - \mathrm{C_2H_4N}], \ \ 104 \ \ (8) \\ [M^+ - \mathrm{C_8H_{16}} - \mathrm{C_2H_3N_2}], \ 78 \ (25) \ [\mathrm{C_5H_4N^+}]. \end{array}$

2-(1-Octyl-5-phenylpyrazol-3-yl)pyridine (**2**c): ¹H NMR (400.13 MHz, 25 °C, CDCl₃): $\delta = 8.56$ (d, ³ $J_{10, 11} = 5.0$ Hz, 11-H), 7.88 (d, ³ $J_{8,9} = 8.0$ Hz, 8-H), 7.65 (t, ³ $J_{9, 10} = 8.1$ Hz, 9-H), 7.39 (m, 5H, H_{Ph}), 7.11 (dd, 10-H), 6.83 (s, 4-H), 4.09 (t, ³ $J_{H,H} = 7.8$ Hz, NCH₂), 1.75 (br, NCH₂CH₂), 1.19 (br, NCH₂CH₂CH₂), 1.11 (br, 8H, CH₂), 0.79 (t, ³ $J_{H,H} = 7.0$ Hz, CH₃); ¹³C{¹H} NMR (100.63 MHz, 25 °C, CDCl₃): $\delta = 152.1$ (C-7), 150.3 (C-3), 149.3 (C-11), 136.1 (C-9), 130.6 (C-5), 128.7 - 128.2 (C_{Pb}), 122.1 (C-10), 119.9 (C-8), 104.4 (C-4), 49.7 (NCH₂), 31.5 - 22.4 (CH₂), 13.9 (CH₃); MS (EI, 70 eV): m/z (%) = 333 (35) [M^-], 304 (9) [$M^+ - C_{gH_3}$], 290 (29) [$M^+ - C_{gH_12}$], 248 (34) [$M^+ - C_{6}$ H₁₃], 235 (52) [$M^+ - C_{8}$ H₁₆], 78 (14) [C_{8} H₄N⁺].

2-(1-Octyl-5-trifluormethyl-3-pyrazolyl)pyridine (2d): ¹H NMR (400.13 MHz, 25 °C, CDCl₃): $\delta = 8.52$ (d, ³ $J_{10,11} = 4.5$ Hz, 11-H), 7.84 (d, ³ $J_{8,9} = 8.0$ Hz, 8-H), 7.65 (dt, ³ $J_{9,10} = 7.8$ Hz, ⁴ $J_{9,11} = 1.5$ Hz, 9-H), 6.15 (s, 4-H), 7.09 (dd, 10-H), 4.15 (t, ³ $J_{H,H} = 7.5$ Hz, NCH₂), 1.84 (br, NCH₂CH₂), 1.23 (br, NCH₂CH₂), 1.18 (br, 8 H, CH₂), 0.77 (t, ³ $J_{H,H} = 7.2$ Hz, CH₃); ¹³C{¹H} NMR (100.63 MHz, 25 °C, CDCl₃): $\delta = 151.0$ (C-7), 150.5 (C-3), 149.3 (C-11), 136.3 (C-9), 133.7 (q, ² $J_{C,F} = 39.0$ Hz, C-5), 122.6 (C-10), 119.9 (q, ¹ $J_{C,F} = 267.7$ Hz, CF₃), 119.8 (C-8), 105.7 (q, ³ $J_{C,F} = 2.0$ Hz, C-4), 51.4 (NCH₂), 31.7–22.4 (CH₂), 13.8 (CH₃); MS (EI, 70 eV): m/z (%) = 325 (13) [M^+], 306 (6) [$M^+ - F$], 282 (25) [$M^+ - C_3H_7$], 268 (24) [$M^+ - F_3$], 256 (84) [$M^+ - C_8H_{15}F_3$], 78 (25) [$C_5H_4N^+$], 51 (8) [$C_4H_3^+$].

2-(5-Methyl-1-octylpyrazol-3-yl)pyrazine (2e): ¹H NMR (400.13 MHz, 25 °C, CDCl₃): δ = 9.03 (d, ⁴*J*_{8,10} = 2.5 Hz, 8-H), 8.39 (d, ³*J*_{10,11} = 3.2 Hz, 11-H), 8.28 (dd, 10-H), 6.51 (s, 4-H), 3.96 (t, ³*J*_{H,H} = 7.5 Hz, NCH₂), 2.19 (s, C5-CH₃), 1.73 (br, NCH₂CH₂), 1.20–1.14 (br, 10H, CH₂), 0.75 (t, ³*J*_{H,H} = 6.5 Hz, CH₂CH₃); ¹³C{¹H} NMR (100.63 MHz, 25 °C, CDCl₃): δ = 148.0 (C-7), 147.2 (C-3), 143.6, 142.3, 141.8 (C-11, C-10, C-8), 139.3 (C-5), 104.1 (C-4), 49.2 (NCH₂), 31.5–22.3 (CH₂), 13.8 (CH₂CH₃), 11.0 (C5–CH₃); MS (EI, 70 eV): *m/z* (%) = 272 (32) [*M*⁺], 257 (4) [*M*⁺ – CH₃], 243 (5) [*M*⁺ – C₂H₅], 229 (16) [*M*⁺ – C₃H₇], 215 (15) [*M*⁺ – C₄H₉], 201 (3) [*M*⁺ – C₅H₁₁], 188 (6) [*M*⁺ – C₆H₁₂], 187 (7) [*M*⁺ – C₆H₁₃], 174 (38) [*M*⁺ – C₇H₁₄], 173 (100) [*M*⁺ – C₈H₁₆ – C₂H₄N₂], 79 (25) [C₄H₃N₂⁺].

2-(4-Chloro-1-octylpyrazol-3-yl)pyridine (2f): Compound 2a (2.57 g, 10.0 mmol) was dissolved in 6% HCl (100 mL) and stirred at room temperature while a saturated solution of chlorine in water (400 mL) was added in small portions over a period of 3 h. After another 2 h, ammonium acetate (15 g) was added and the pale yellow solution was neutralized with KOH. The oily product was extracted with ethyl acetate (2×25 mL) and dichloromethane (1 × 10 mL). The organic phase was washed with water (2 × 25 mL), dried over MgSO₄ and the solvent was removed in vacuo. Yield: 2.86 g (98%) colorless oil. ¹H NMR (400.13 MHz, 25 °C, CDCl₃): $\delta = 8.68$ (d, ${}^{3}J_{10,11} = 4.3$ Hz, 11-H), 7.91 (d, ${}^{3}J_{8,9} = 7.9$ Hz, 8-H), 7.68 (t, ${}^{3}J_{9,10} = 7.9$ Hz, 9-H), 7.42 (s, 5-H), 7.19 (dd, 10-H), 4.07 (t, ${}^{3}J = 7.3$ Hz, NCH2CH2), 1.82 (quint, NCH2CH2), 1.28-1.20 (m, 10H, CH2), 0.80 (t, ${}^{3}J = \tilde{6}.7 \text{ Hz}, \text{ CH}_{3}$; ${}^{13}\text{C}{}^{1}\text{H}$ NMR (100.63 MHz, 25 °C, CDCl₃): $\delta = 152.0$ (C-7), 150.6 (C-11), 149.8 (C-3), 136.5 (C-9), 128.9 (C-5), 122.4 (C-10), 122.0 (C-8), 108.1 (C-4), 53.1 (NCH₂), 31.6-22.4 (CH₂), 13.9 (CH₃); MS (EI, 70 eV, ³⁵Cl): m/z (%) = 291 (33) $[M^+]$, 276 (2) $[M^+ - CH_3]$, 262 (8) $[M^+ - C_2H_5], 248 (20) [M^+ - C_3H_7], 234 (15) [M^+ - C_4H_6], 220 (14)$ $\begin{bmatrix} M^{+} - C_{5}H_{11} \end{bmatrix}, 206 (39) \begin{bmatrix} M^{+} - C_{6}H_{13} \end{bmatrix}, 192 (40) \begin{bmatrix} M^{+} - C_{7}H_{15} \end{bmatrix}, 179 (58) \\ \begin{bmatrix} M^{+} - C_{8}H_{16} \end{bmatrix}, 158 (5) \begin{bmatrix} M^{+} - C_{7}H_{14} - C \end{bmatrix}, 144 (8) \begin{bmatrix} M^{+} - C_{8}H_{16} - C \end{bmatrix},$ 130 (21) $[M^+ - C_8H_{16}N - Cl]$, 117 (12) $[M^+ - C_8H_{15}N_2 - Cl]$, 78 (30) $[C_5H_4N^+].$

2-(4-Bromo-1-octylpyrazol-3-yl)pyridine (2g): Compound 2a (2.57 g, 10.0 mmol) was dissolved in 20% acetic acid (100 mL) and stirred at room temperature while bromine (1.59 g, 10.0 mmol) dissolved in glacial acetic acid (10 mL) was added over 15 min. The resulting pale yellow solution was neutralized with KOH. The oily product was extracted with ethyl acetate

 $\begin{array}{ll} (2\times25\ {\rm mL}). \ {\rm The\ organic\ phase\ was\ washed\ with\ water\ (2\times25\ {\rm mL})\ {\rm and\ dried\ over\ MgSO_4,\ {\rm and\ the\ solvent\ was\ removed\ in\ vacuo.\ Yield:\ 2.46\ g\ (73\ \%)\ pale\ yellow\ oil.\ ^1{\rm H\ NMR}\ (400.13\ {\rm MHz},\ 25\ ^{\circ}{\rm C},\ {\rm CDCl}_3):\ \delta=8.68\ (ddd,\ ^3\ J_{10,11}=4.2\ {\rm Hz},\ ^4\ J_{0,11}=1.8\ {\rm Hz},\ ^5\ J_{8,11}=0.9\ {\rm Hz},\ 11-{\rm H}).\ 7.92\ (dt,\ ^3\ J_{8,9}=7.9\ {\rm Hz},\ ^4\ J_{9,11}=1.8\ {\rm Hz},\ ^5\ J_{8,11}=0.9\ {\rm Hz},\ 11-{\rm H}).\ 7.92\ (dt,\ ^3\ J_{8,9}=7.9\ {\rm Hz},\ ^4\ J_{8,10}=0.9\ {\rm Hz},\ 8+{\rm H}),\ 7.69\ (dt,\ ^3\ J_{9,10}=7.9\ {\rm Hz},\ 9+{\rm H}),\ 7.45\ (s,\ 5-{\rm H}),\ 7.19\ (ddd,\ 10-{\rm H}),\ 4.10\ (t,\ ^3\ J=7.3\ {\rm Hz},\ {\rm NCH}_2{\rm CH}_2).\ 1.89\ ({\rm quint},\ {\rm NCH}_2{\rm CH}_2),\ 1.35\ 1.20\ (m,\ 10\,{\rm H},\ {\rm CH}_2),\ 0.82\ (t,\ ^3\ J=6.9\ {\rm Hz},\ {\rm CH}_3);\ 1^3{\rm C}\{^1{\rm H}\}\ {\rm NMR}\ (100.63\ {\rm MHz},\ 25\ ^{\circ}{\rm C},\ {\rm CDCl}_3);\ \delta=149.9\ ({\rm C}-7),\ 148.4\ ({\rm C}-11),\ 146.1\ ({\rm C}-3),\ 135.2\ ({\rm C}-9),\ 130.1\ ({\rm C}-5),\ 121.4\ ({\rm C}-10),\ 121.0\ ({\rm C}-8),\ 90.6\ ({\rm C}-4),\ 52.1\ ({\rm NCH}_2),\ 30.5-19.7\ ({\rm CH}_2),\ 12.9\ ({\rm CH}_3);\ {\rm MS}\ ({\rm EI},\ 70\ {\rm eV},\ ^{79}{\rm Br});\ m/z\ (\%) = 335\ (36)\ [M^+],\ 320\ (5)\ [M^+-{\rm C}_4{\rm H}_3],\ 306\ (24)\ [M^+-{\rm C}_2{\rm H}_3],\ 292\ (66)\ [M^+-{\rm C}_3{\rm H}_1],\ 250\ (44)\ [M^+-{\rm C}_8{\rm H}_{16}],\ 293\ (5)\ [M^+-{\rm C}_8{\rm H}_{16}],\ 209\ (5)\ [M^+-{\rm C}_8{\rm H}_{16}],\ 200\ (5)\ [M^$

2-(4-Nitro-1-octylpyrazol-3-yl)pyridine (2h): Compound 2a (2.57 g, 10.0 mmol) was dissolved in 80% H₂SO₄ (10 mL) at 0 °C. To this solution, a mixture of 65 % HNO3 (5 mL) and 80 % H_2SO4 (5 mL) was added dropwise over a period of 30 min. After the reaction mixture had been heated to 90 °C for 4 h, it was poured over ice (300 g) and carefully neutralized with conc. K_2CO_3 . The oily product was extracted with ethyl acetate (2 × 25 mL) and dichloromethane (1 × 10 mL), the organic phase was washed with water $(2 \times 25 \text{ mL})$, dried over MgSO₄ and the solvent was removed in vacuo. Yield: 2.12 g (70%), yellow oil. ¹H NMR (400.13 MHz, 25 °C, CDCl₃): $\delta = 8.66$ (d, ${}^{3}J_{10,11} = 4.5$ Hz, 11-H), 8.19 (s, 5-H), 7.72 (dt, ${}^{3}J_{8,9} = {}^{3}J_{9,10} = 7.5$ Hz, ${}^{4}J_{9,11} = 1.5$ Hz, 9-H), 7.66 (d, 8-H), 7.19 (ddd, ${}^{4}J_{8,10} = 1.5$ Hz, 10-H), 4.11 (t, ${}^{3}J = 7.3 \text{ Hz}$, NCH₂CH₂), 1.86 (quint, NCH₂CH₂), 1.25-1.15 (m, 10 H, CH_2), 0.79 (t, ${}^{3}J = 6.9 Hz$, CH_3); ${}^{13}C{}^{1}H{}$ NMR (100.63 MHz, 25°C, CD-Cl₃): $\delta = 149.4$ (C-11), 149.0 (C-7), 146.0 (C-3), 136.1 (C-9), 132.7 (C-4), 130.2 (C-5), 124.7 (C-10), 123.7 (C-8), 53.5 (NCH₂), 31.5-22.3 (CH₂), 13.8 (CH_3) ; MS (EI, 70 eV): m/z (%) = 302 (5) $[M^+]$, 301 (5) $[M^- - H]$, 287 (12) $[M^+ - CH_3]$, 285 (100) $[M^+ - OH]$, 273 (29) $[M^+ - C_2H_5]$, 259 (29) $[M^+ - C_3H_7]$, 245 (22) $[M^+ - C_4H_9]$, 231 (18) $[M^+ - C_5H_{11}]$, 217 (39) $[M^+ - C_6 H_{13}], 203 (64) [M^+ - C_7 H_{15}], 190 (29) [M^+ - C_8 H_{16}], 105 (30)$ $[C_6H_5N^+]$, 78 (41) $[C_5H_4N^+]$.

Syntheses of the peroxo complexes 3b-h:

Method A: A solution of an octyl-substituted pyrazolylpyridine (2b-h, 3.00 mmol) in methanol (30 mL) was added to a solution of molybdic acid (0.97 g, 6.00 mmol) in 30 % H₂O₂ (30 mL), and the resulting mixtures were vigorously stirred for 10 min. The complexes were extracted with CH₂Cl₂ (2 × 50 mL). To remove traces of H₂O₂, the combined organic solutions were washed with water (2 × 50 mL) and dried over MgSO₄. Evaporation of the solvent yielded yellow solids, which were washed with Et₂O (2 × 30 mL) to remove organic impurities.

Method B: A solution of an octyl-substituted pyrazolylpyridine (2b-h, 3.00 mmol) in CH₂Cl₂ (20 mL) was added to a solution of [MoO- $(O_2)_2 \cdot (DMF)_2$] (1.93 g, 6.00 mmol) in CH₂Cl₂ (30 mL) and the resulting mixture was stirred for 1 h. The solvent was evaporated to dryness and the resulting yellow solids were extracted with diethyl ether (2 × 20 mL) to remove organic impurities. Extraction with hot ethyl acetate and evaporation of the yellow solutions yielded the desired peroxo complexes. Isolated yields of peroxo complexes: 60–80%, yellow microcrystalline solids.

[2-(5-Methyl-1-octylpyrazol-3-yl)pyridine]oxodiperoxomolybdenum (vi) (3b): IR (KBr): $\tilde{\nu} = 3134 \text{ m cm}^{-1}$, 3086 w, 3059 w, 2955 s, 2917 vs. 2868 s. 2851 s, 1611 s, 1567 w, 1517 m, 1467 s, 1459 s, 1443 s, 1424 s, 1379 s. 1327 m, 1290 w, 1252 w, 1217 m, 1160 m, 1111 m, 1054 m, 1025 w, 949 vs (Mo=O). 878 s. 866 vs (2 × O-O). 820 m, 788 vs, 752 w, 658 s, 643 m, 585 s, 538 m; ⁻¹H NMR (400.13 MHz, 25 · C, CDCl₃): isomer A: $\delta = 9.16$ (d, ${}^{3}J_{10,11} = 6.0$ Hz, 11-H). 8.17 (t, ${}^{3}J_{8,9} = {}^{3}J_{9,10} = 6.4$ Hz, 9-H), 7.86 (d, 8-H), 7.58 (t, 10-H). 6.46 (s, 4-H), 3.98 (t, ${}^{3}J_{11,H} = 8.0$ Hz, NCH₂CH₂). 2.19 (s, C5-CH₃). 1.62 (br, NCH₂CH₂), 1.3-1.1 (m, 10H, CH₂). 0.80 (t, ${}^{3}J_{H,H} = 7.0$ Hz, CH₃); isomer **B**: $\delta = 8.15$ (d, ${}^{3}J_{10,11} = 4.5$ Hz, 11-H), 7.74 (t, ${}^{3}J_{8,9} = {}^{3}J_{9,10} = 7.5$ Hz, 9-H), 7.59 (d, 8-H), 7.16 (t, 10-H). 6.82 (s, 4-H), 4.55 (t, ${}^{3}J_{H,H} = 7.5$ Hz, NCH₂), 2.53 (s, C5-CH₃), 1.97 (br, NCH₂CH₂), 1.3-1.1 (m, 10H, CH₂), 0.82 (t, ${}^{3}J_{H,H} = 7.0$ Hz, CDH₃); isomer ratio A/B: 0.98; ${}^{13}C{}^{1}H$ NMR (100.63 MHz, 25 °C, CDCl₃): isomer A: $\delta = 154.3$ (C-11), 151.1 (C-7), 146.8 (C-3), 143.8 (C-5), 142.8 (C-9), 125.8 (C-10), 122.4 (C-8), 103.8 (C-4), 49.5 (NCH₂), 31.6-22.4 (CH₂), 14.0 (CH₃), 11.1 (C5-CH₃); isomer **B**: $\delta = 151.1$ $\begin{array}{l} (C\text{-7}),\,147.4\,(C\text{-3}),\,146.8\,(C\text{-11}),\,145.0\,(C\text{-5}),\,139.0\,(C\text{-9}),\,124.8\,(C\text{-10}),\,120.4\\ (C\text{-8}),\,105.2\,(C\text{-4}),\,49.5\,(\text{NCH}_2),\,31.6\text{--}22.4\,(\text{CH}_2),\,14.0\,(\text{CH}_3),\,12.3\,(\text{C}5\text{-}C\text{H}_3);\,C_{17}\text{H}_{23}\text{MoN}_3\text{O}_5\,(447.4)\text{: calcd C}\,45.64,\,\text{H}\,5.63,\,\text{N}\,9.39\text{; found C}\,45.42,\,\text{H}\,5.51,\,\text{N}\,9.28. \end{array}$

[2-(1-Octyl-5-phenylpyrazol-3-yl)pyridine]oxodiperoxomolybdenum (vi) (3c): IR (KBr): $\tilde{v} = 3124 \text{ w cm}^{-1}$, 3094 w, 3062 w, 3031 w, 2953 s, 2924 vs, 2854 s, 1613s, 1567m, 1521m, 1483m, 1460s, 1438s, 1379m, 1341w, 1294w, 1250w, 1160 w, 1112 m, 1062 m, 1027 w, 1015 m, 974 m, 952 vs (Mo=O), 876 s, 864 vs $(2 \times O - O)$, 784s, 765s, 700s, 678m, 663s, 584s, 535m; ¹H NMR $(400.13 \text{ MHz}, 25 \degree \text{C}, \text{CDCl}_3)$: isomer A: $\delta = 9.19 (\text{d}, {}^{3}J_{10,11} = 5.5 \text{ Hz}, 11\text{-H})$, 8.20 (t, ${}^{3}J_{8,9} = {}^{3}J_{9,10} = 7.0$ Hz, 9-H), 7.93 (d, 8-H), 7.59 (t, 10-H), 6.63 (s, 4-H), 4.06 (t, ${}^{3}J_{\text{H, H}} = 7.8 \text{ Hz}$, NCH₂CH₂), 1.48 (br, NCH₂CH₂), 1.2-1.0 (m, 10 H, CH₂), 0.75 (t, ${}^{3}J_{H, H} = 7.8$ Hz, CH₃); isomer **B**: $\delta = 8.18$ (d, ${}^{3}J_{10,11} = 3.0$ Hz, 11-H), 7.76 (t, ${}^{3}J_{8,9} = {}^{3}J_{9,10} = 7.8$ Hz, 9-H), 7.63 (d, 8-H), 3 _{10,11} 10 10 10 11 10 10 11 10 11 10 11 10 11 10 11 11 10 11 11 10 11 11 10 11 NCH₂CH₂), 1.2-1.0 (m, 10 H, CH₂), 0.76 (t, ${}^{3}J_{H,H} = 7.8$ Hz, CH₃); isomer ratio A/B: 1.42; ¹³C{¹H} NMR (100.63 MHz, 25 °C, CDCl₃): isomer A: $\delta = 154.4$ (C-11), 150.9 (C-7), 146.6 (C-3), 144.3 (C-5), 142.8 (C-9), 131.7 127.7 (C_{Pb}), 125.0 (C-10), 122.6 (C-8), 104.5 (C-4), 50.0 (NCH₂), 31.5-22.4 (CH_2) , 14.0 (CH_3) ; isomer **B**: $\delta = 151.7$ (C-3, C-7), 147.0 (C-11), 144.9 (C-5), 139.1 (C-9), 131.7-127.7 (C_{Ph}), 125.0 (C-10), 120.5 (C-8), 105.5 (C-4), 50.0 (NCH₂), 31.5–22.4 (CH₂), 14.0 (CH₃); $C_{22}H_{27}MoN_3O_5$ (509.4): calcd C 51.87, H 5.34, N 8.25; found C 51.64, H 5.18, N 8.17.

2-(1-Octyl-5-trifluoromethyl-3-pyrazolyl)pyridineoxodiperoxomolybdenum(vi) (3d): IR (KBr): $\tilde{v} = 3121 \,\mathrm{m}\,\mathrm{cm}^{-1}$, 3092 w, 2954 m, 2927 s, 2858 m, 1612 m, 1558 w, 1442 m, 1417 w, 1383 w, 1347 m, 1275 s, 1217 m, 1174 vs, 1141 vs (2×CF), 1087 w, 1061 m, 1041 w, 1028 w, 963 s (Mo=O), 880 m, 870 s (2×O-O). 842w, 788s, 750w, 729w, 696w, 668m, 592m, 544w; ¹HNMR $(400.13 \text{ MHz}, 25 \degree \text{C}, \text{CDCl}_3)$: isomer A: $\delta = 9.28 \text{ (d, }^3J_{10, 11} = 5.5 \text{ Hz}, 11\text{-H})$, 8.30 (dt, ${}^{3}J_{8,9} = {}^{3}J_{10,9} = 8.0$ Hz, ${}^{4}J_{9,11} = 1.5$ Hz, 9-H), 7.99 (d, 8-H), 7.72 (dd, 10-H), 7.05 (s, 4-H), 4.21 (t, ${}^{3}J_{H,H} = 8.3$ Hz, NCH₂), 1.75 (m, NCH₂CH₂), 1.4–1.3 (m, 10H, CH₂). 0.85 (t, ${}^{3}J_{H,H} = 6.3$ Hz, CH₂); isomer B: $\delta = 8.25$ (d, ${}^{3}J_{10,11} = 4.0$ Hz, 11-H), 7.86 (t, ${}^{3}J_{8,9} = {}^{3}J_{10,9} = 8.2$ Hz, 9-H), 7.70 (d, 8-H), 7.32 (s, 4-H), 7.28 (dd, 10-H), 4.78 (t, ${}^{3}J_{H,H} = 7.3$ Hz, NCH2), 2.12 (m, NCH2CH2), 1.3-1.2 (m, 10H, CH2), 0.82 (t, ${}^{3}J_{\text{H,H}} = 6.3 \text{ Hz}, \text{CH}_{3}$; isomer ratio A/B: 28.9; ${}^{13}\text{C}\{{}^{1}\text{H}\}$ NMR (100.63 MHz, 25 °C, CDCl₃): isomer A: $\delta = 155.0$ (C-11), 149.7 (C-7), 144.2 (C-3), 143.2 (C-9), 134.5 (q, ${}^{2}J_{C, F} = 40.8 \text{ Hz}$, C-5), 125.9 (C-10), 123.0 (C-8), 118.6 (q, ${}^{1}J_{C,F} = 270.2 \text{ Hz}, \text{ CF}_{3}$, 105.4 (q, ${}^{3}J_{C,F} = 3.1 \text{ Hz}, \text{ C-4}$), 52.4 (NCH₂), 31.7– 22.6 (CH₂), 14.0 (CH₃); isomer **B**: no signals identified; ¹⁹F{¹H} NMR $(376.48 \text{ MHz}, 25 \degree \text{C}, \text{CDCl}_3)$: isomer A: $\delta = 2.35 (\text{s}, \text{CF}_3)$, isomer B: $\delta = 2.05$ (s, CF₃), isomer ratio A/B: 23.4; C₁₇H₂₂F₃MoN₃O₅ (501.3): calcd C 40.73, H 4.42, N 8.38; found C 39.67, H 4.35, N 8.11.

2-(5-Methyl-1-octylpyrazol-3-yl)pyrazineoxodiperoxomolybdenum(vi) (3e): IR (KBr): $\tilde{v} = 3122 \text{ w cm}^{-1}$, 3074 w, 2955 s, 2926 vs, 2854 s, 1532 m, 1458 s, 1426 s, 1406 s, 1367 m, 1226 w, 1172 m, 1162 s, 1110 w, 1108 w, 1058 m, 1044 s, 959 vs, 944 vs $(2 \times Mo=O)$, 868 vs (O-O), 825 w, 662 m, 644 w, 586 s, 542 m; ¹H NMR (400.13 MHz, 25 °C, CDCl₃): isomer A: $\delta = 9.23$ (d, ${}^{4}J_{8,10} = 1.0$ Hz, 8-H), 9.12 (dd, ${}^{3}J_{10,11} = 3.0$ Hz, 10-H), 8.94 (d, 11-H), 6.60 (s, 4-H), 3.96 (t, ${}^{3}J_{H,H} = 8.0$ Hz, NCH₂), 2.24 (s, C5-CH₃), 1.99 (br, NCH₂CH₂), 1.30–1.13 (br, 10 H, CH₂), 0.80 (t, ${}^{3}J_{H, H} = 6.5$ Hz, CH₂CH₃); isomer **B**: $\delta = 8.92$ (d, ${}^{4}J_{8,10} = 1.5$ Hz, 8-H), 8.52 (d, ${}^{3}J_{10,11} = 2.5$ Hz, 11-H). 8.15 (dd, 10-H), 6.95 (s, 4-H), 4.59 (t, ${}^{3}J_{H,H} = 7.5$ Hz, NCH₂), 2.58 (s, C5-CH₃), 1.99 (br, NCH₂CH₂), 1.30-1.13 (br, 10H, CH₂), 0.80 (t, ${}^{3}J_{\text{H.H}} = 6.5 \text{ Hz}, \text{ CH}_{2}\text{CH}_{3}$; isomer ratio A/B: 0.084; ${}^{13}\text{C}\{{}^{1}\text{H}\}$ NMR $(100.63 \text{ MHz}, 25 \degree \text{C}, \text{CDCl}_3)$: isomer A: $\delta = 146.2, 145.8, 145.2, 144.5, 142.6,$ 141.2 (C-3, C-5, C-7, C-8, C-10, C-11), 104.4 (C-4), 49.9 (NCH₂), 31.6-22.5 (CH₂), 13.9 (CH₂CH₃), 11.2 (C5-CH₃); isomer **B**: $\delta = 148.5$ (C-3), 147.8 (C-7), 146.2, 141.2, 140.1 (C-11, C-10, C-8), 140.3 (C-5), 105.7 (C-4), 49.8 (NCH₂), 31.6 22.5 (CH₂), 13.9 (CH₂CH₃), 12.4 (C5-CH₃); C16H24MoN4O5 (448.34): calcd C 42.86, H 5.40, N 12.50; found C 42.37, H 5.41, N 12.53.

2-(4-Chloro-1-octyl-3-pyrazolyl)pyridineoxodiperoxomolybdenum(v1) (3f): IR (KBr): $\tilde{v} = 3132 \text{ m cm}^{-1}$, 3120 m, 2954s, 2924 vs, 2854s, 1611s, 1566 w, 1522 m, 1467 m, 1435 s, 1373 m, 1334 m, 1290 w, 1251 w, 1217 m, 1166 m, 1122 w, 1109 w, 1065 m, 1056 m, 1032 w, 1002 m (C-Cl), 956 vs, 947 vs (2 × Mo=O), 878 s, 865 vs (2 × O - O), 881 m, 790 s, 754 m, 690 w. 666 m, 635 m, 588 s, 534 m; ¹H NMR (400.13 MHz, 25 °C, CDCl₃): isomer A: $\delta = 9.35$ (d, ${}^{3}J_{10,11} = 5.5$ Hz, 11-H), 8.59 (d, ${}^{3}J_{8,10} = 1.5$ Hz, 10-H), 7.38

(s, 5-H), 4.14 (t, ${}^{3}J_{H,H} = 7.3$ Hz, NCH₂CH₂), 1.73 (br, NCH₂CH₂), 1.30–1.20 (m, 10 H, CH₂), 0.81 (t, ${}^{3}J_{H,H} = 6.9$ Hz, CH₃); isomer **B**: $\delta = 8.41$ (d, ${}^{3}J_{10,11} = 4.0$ Hz, 11-H), 8.26 (br, 8-H), 8.05 (s, 5-H), 7.86 (dt, ${}^{3}J_{8.9} = {}^{3}J_{9,10} = 7.8$ Hz, ${}^{4}J_{9,11} = 1.5$ Hz, 9-H), 7.30 (dd, 10-H), 4.69 (t, ${}^{3}J_{H,H} = 7.3$ Hz, NCH₂CH₂), 2.06 (br, 2H, NCH₂CH₂), 1.30–1.20 (m, 10 H, CH₂), 0.81 (t, ${}^{3}J_{H,H} = 6.9$ Hz, 3H, CH₃); isomer ratio **A**/**B**: 7.05; 13 C{¹H} NMR (100.63 MHz, 25 °C, CDCl₃): isomer **A**: $\delta = 155.1$ (C-11), 149.8 (C-7), 142.9 (C-9), 149.6 (C-3), 130.8 (C-5), 125.5 (C-10), 123.4 (C-8), 190.7 (C-4), 53.06 (NCH₂), 32.0–22.5 (CH₂), 14.03 (CH₃); isomer **B**: $\delta = 155.0$ (C-3), 147.5 (C-11), 143.8 (C-7), 139.2 (C-9), 136.6 (C-5), 123.4 (C-10), 121.5 (C-8), 110.6 (C-4), 53.97 (NCH₂), 32.0–21.5 (CH₂), 14.03 (CH₃); C₁₆H₂₂CIMON₃O₅ (467.78): calcd C 41.08, H 4.74, N 8.98; found C 40.64, H 4.92, N 8.91.

2-(4-Bromo-1-octyl-3-pyrazolyl)pyridineoxodiperoxomolybdenum(vi) (3g): IR (KBr): $\tilde{v} = 3103 \,\mathrm{s} \,\mathrm{cm}^{-1}$, 2952 s, 2923 vs, 2853 s, 1612 s, 1594 w, 1568 w, 1522 w, 1506 w, 1464 m, 1436 m, 1430 m, 1370 w, 1334 w, 1261 w, 1251 w, 1207 w, 1156 w, 1108 w, 1068 w, 1056 w, 1031 w, 992 w (C-Br), 954 vs (Mo=O), 878 m, 863 vs (2×O-O), 788 m, 751 w, 690 w, 666 m, 634 w, 587 m, 541 w; ¹H NMR (400.13 MHz, 25 °C, CDCl₃): isomer A: $\delta = 9.30$ (dd, ${}^{3}J_{10,11} = 5.5 \text{ Hz}, {}^{4}J_{9,11} = 5.5 \text{ Hz}, 11\text{-H}), 8.69 \text{ (d, } {}^{3}J_{8,9} = 7.6 \text{ Hz}, 8\text{-H}), 8.25 \text{ Hz}, 11\text{-H})$ (dt, ${}^{3}J_{9,10} = 7.5$ Hz, 9-H), 7.68 (ddd, ${}^{4}J_{8,10} = 1.5$ Hz, 10-H), 7.37 (s, 5-H), 4.10 (t, ${}^{3}J_{11,H} = 7.3$ Hz, NCH₂CH₂), 1.69 (br, NCH₂CH₂), 1.30–1.20 (m, 10 H, CH₂), 0.81 (t, ${}^{3}J_{H,H} = 6.9$ Hz, CH₃); isomer **B**: $\delta = 8.38$ (d, ${}^{3}J_{10,11} = 8.0$ Hz, 11-H), 8.03 (s, 5-H), 7.92 (d, ${}^{3}J_{8,9} = 7.9$ Hz, 8-H), 7.82 (dt, ${}^{3}J_{9,10} = 7.9$ Hz, 9-H), 7.69 (dd, 10-H), 4.66 (t, ${}^{3}J_{\text{H,H}} = 7.3$ Hz, NCH₂CH₂), 2.01 (br, NCH₂CH₂), 1.30–1.20 (m, 10H, CH₂), 0.81 (t, ${}^{3}J_{\text{II,H}} = 6.9$ Hz, CH₃); isomer ratio **Å**/**B**: 7.86; ¹³C{¹H} NMR (100.63 MHz, 25 °C, CDCl₃): isomer A: $\delta = 155.1$ (C-11), 150.1 (C-7), 144.1 (C-9), 140.9 (C-3), 133.2 (C-5), 125.5 (C-10), 123.2 (C-8), 92.82 (C-4), 53.00 (NCH₂), 32.0-21.5 (CH₂), 14.03 (CH₃); isomer **B**: δ = 155.0 (C-3), 148.9 (C-7), 147.5 (C-11), 144.1 (C-9), 139.1 (C-5), 122.6 (C-10), 121.4 (C-8), 93.64 (C-4), 53.93 (NCH2), 32.0-21.5 (CH₂), 14.03 (CH₃); C₁₆H₂₂BrMoN₃O₅ (512.2): calcd C 37.52, H 4.33, N 8.20; found C 37.98, H 4.37, N 8.40.

2-(4-Nitro-1-octyl-3-pyrazoly])pyridineoxodiperoxomolybdenum(vi) (**3h**): IR (KBr): $\tilde{\nu} = 3139 \,\mathrm{m}\,\mathrm{cm}^{-1}$, 3101 m, 2953 s, 2926 vs, 2855 s, 1619 m, 1570 m, 1540 vs, 1504 vs (N=O)_{as}, 1453 s, 1442 s, 1369 s, 1350 vs (N=O)_{sym}, 1224 m, 1147 w, 1112 w, 1038 w, 957 vs (Mo=O), 875 m, 864 vs (2 × O-O), 835 s, 790 m, 755 s, 666 m, 635 w, 617 w, 585 s, 536 m; ¹H NMR (400.13 MHz, 25 °C, CDCl₃): isomer A: $\delta = 9.43$ (br, 11-H), 9.10 (d, ${}^{3}J_{8,9} = 8.0$ Hz, 8-H), 8.39 (t, ${}^{3}J_{9,10} = 8.0$ Hz, 9-H), 8.23 (s, 5-H), 7.86 (t, ${}^{4}J_{10,11} = 6.3$ Hz, 10-H), 4.23 (t, ${}^{3}J_{\mathrm{H,H}} = 7.0$ Hz, NCH₂CH₂), 1.78 (br, NCH₂CH₂), 1.31 - 1.20 (m, 10 H, CH₂), 0.84 (t, ${}^{3}J_{\mathrm{H,H}} = 7.0$ Hz, CH₃); isomer B: δ not detectable; isomer ratio A/B: >50; ${}^{13}C{}^{14}$ NMR (100.63 MHz, 25 °C, CDCl₃): isomer A: $\delta = 155.6$ (C-11), 147.5 (C-7), 147.5 (C-3), 143.4 (C-9), 138.6 (C-5), 133.9 (C-4), 127.4 (C-10), 127.1 (C-8), 53.6 (NCH₂), 31.6-22.5 (CH₂), 14.0 (CH₃); isomer B: δ not detectable; c₁₆H₂₂MoN₄O₇ (478.31): calcd C 40.18, H 4.64, N 11.71: found C 39.07, H 4.59, N 11.59.

Catalytic epoxidation of cyclooctene: The reactions were carried out in a two-neck 100 mL flask equipped with a reflux condenser and a Quickfit^{*} septum adapter. In this flask cyclooctene (1.00 g, 9.07 mmol), dibutyl ether (1.00 g, 7.75 mmol, internal standard) and *t*BuOOH (1.35 mL of a 6.90 M solution in CHCl₃) were dissolved in CHCl₃ (23 mL). The solution was heated to reflux temperature (61 °C, oil bath temperature: 95 °C) and the catalysts (12–13 mg) dissolved in CHCl₃ (2 mL) were added from a PE syringe. Samples were taken after 2, 5, 10, 20, and 40 min with a PE syringe.

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